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RESEARCH ARTICLE

EPITHELIAL DYSPLASIAS AND THEIR HORMONE RECEPTOR EXPRESSIONS IN HUMAN UTERINE CERVIX CAUSED BY HUMAN PAPILLOMA VIRUS

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ABSTRACT

Cervical cancer is a major cause of morbidity and mortality in developing countries like India. The pre-neoplastic stages of cervical cancer are Low Grade Squamous Intraepithelial Lesion (LSIL) and High Grade Squamous Intraepithelial Lesion (HSIL). The major aetiology is high-risk human papilloma virus infection by subtypes like HPV 16,18,33 etc. The human uterine cervical epithelium responds to the sex hormones like oestrogen and progesterone. Many neoplastic and pre-neoplastic conditions of the cervix are known to alter the expression of these receptors. This study aims to evaluate the various degrees of expression of oestrogen and progesterone receptors in epithelial dysplasias of human uterine cervix caused by High Risk Human Papilloma Virus. 80 cases of cervical biopsies were subjected to Polymerase Chain reaction for High Risk Human Papilloma Virus and histological grading. Immunohistochemical staining was done and nuclear positivity for oestrogen and progesterone receptors was assessed, tabulated and statistically analysed. Both receptors showed significant reduction in the expression of ER and PR in increasing grade of epithelial dysplasia and absent in carcinoma. Dysplastic and malignant changes in the epithelium are accompanied by down regulation of the hormone receptors. This might help in the early detection of cervical cancer.

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INTRODUCTION

Cervical cancer can be effectively prevented. Many developed countries have achieved near eradication of this cancer. This is due to the fact that it progresses gradually and easily accessible for examination and screening by PAP smear. The pre-neoplastic stages of cervical cancer are Low Grade Squamous Intraepithelial Lesion (LSIL) and High Grade Squamous Intraepithelial Lesion (HSIL) leading to Squamous Cell Carcinoma (SCC). Still it is a major cause of morbidity and mortality in many developing countries like India. There are many etiological factors for cervical cancer which includes smoking, oral contraceptive use, immune suppression and hormone replacement therapy (Angiolo et al., 2011).

But the major aetiology is high-risk human papilloma virus (HR-HPV) infection by subtypes like HPV 16, 18, 33 etc. The human uterine cervical epithelium responds to the sex hormones like oestrogen and progesterone. This is due to the presence of corresponding Oestrogen Receptor (ER) and Progesterone Receptor (PR) on the nuclear membrane (Remoue et al., 2003). The growth and differentiation of the epithelium is regulated by the hormones via these receptors (Sarpin et al., 2009).

These receptors belong to the nuclear receptor super family of transcription factors. Oestrogen receptor- α is the major receptor in the uterine cervix (Pearce et al., 2004). Many neoplastic and pre-neoplastic conditions of the cervix are known to alter the expression of these receptors (Chung et al., 2013).

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MATERIALS AND METHODS

Study subjects

Inclusion criteria: Women aged 30-60 years were selected.
Exclusion criteria: History of hysterectomy/ conisation/ previous treatment of cervical cancer. This study was approved by the Institutional Human Ethical Committee (IHEC).

Sampling

Punch biopsy of the cervix was taken. It was divided in to two. One part was stored in Phosphate Buffered Saline (PBS) for PCR. The other part was preserved in Neutral Buffered Formalin (NBF) 10% for histopathological studies.

Screening for HPV

The PBS stored cervical tissue sample was used for PCR. HR-HPV was detected using (*AmpliGenei HPV detection kit, Bangalore Genei, India*). The samples which were confirmed for HR-HPV by type-specific PCR (HPV 16, 18, 31, 33, 35, 45, 52 and 58) were separated. The negative cases were used as controls. The total number of patients thus selected was 80.

Histological grading

The cervical biopsies stored in the formalin were processed by the routine histopathology method to obtain paraffin blocks. The sections were stained by Hematoxylin & Eosin method and evaluated microscopically. They were graded to normal, LSIL, HSIL and SCC. Apart from the 20 normal biopsies which acted as controls for hormone receptor estimation there were 20cases of LSIL, HSIL and SCC.

Immunohistochemistry for ER & PR

Immunohistochemistry was done from the paraffin sections using DAKO IHC kit by Labelled Streptavidin-biotin (LSAB) method. For Oestrogen receptor *DAKO Monoclonal rabbit Anti-Human Oestrogen receptor- α antibody clone SP1* and for Progesterone receptor *DAKO Monoclonal mouse Anti-Human Progesterone receptor antibody clone PgR636* were used. The nuclear positivity of both receptors was assessed and the results were tabulated. The results were analysed by Chi-Squared test.

RESULTS

Out of the 80 biopsies selected which include normal, LSIL, HSIL and SCC, the expression of ER and PR was estimated (Figures 2 to 9) and tabulated (Tables 1 & 2).

Table 1. Immunohistochemical expression of Oestrogen Receptor (ER)

Study population (n=80)	Number of positive biopsies for Oestrogen Receptor	Percentage %
Controls (n=20)	17	85
LSIL (n=20)	5	25
HSIL (n=20)	2	10
SCC (n=20)	-	-

Data are expressed as percentage. (Each category n=20).
Chi-squared test: Calculated value 26.12 vs Table value 5.99.

Table 2. Immunohistochemical expression of Progesterone Receptor

Study population (n=80)	Number of positive biopsies for Progesterone receptor	Percentage %
Controls (n=20)	15	75
LSIL (n=20)	8	40
HSIL (n=20)	5	25
SCC (n=20)	-	-

Data are expressed as percentage. (Each category n=20).
Chi-squared test: Calculated value 15.66 vs Table value 5.99).

The loss of receptors in increasing dysplasias can be appreciated in the comparative bar chart (Figure 1).

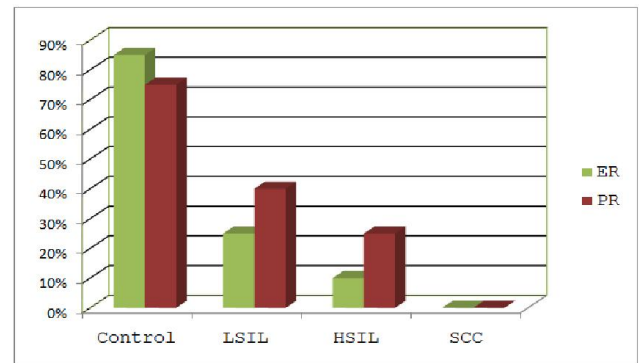


Figure 1. Comparative expression of ER and PR in dysplasias of cervix including SCC

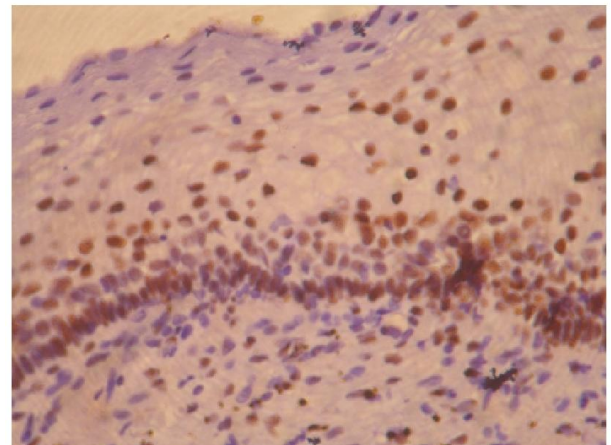


Figure 2. ER expression in normal cervix

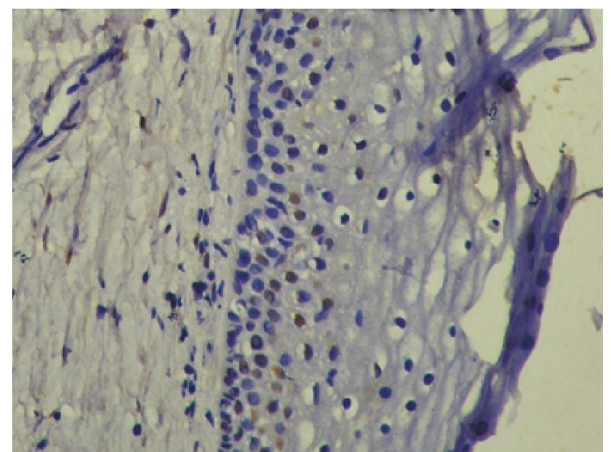


Figure 3. ER expression in LSIL

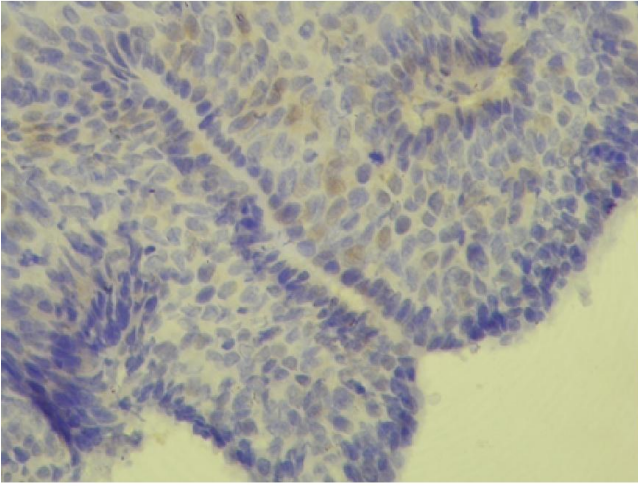


Figure 4. ER expression in HSIL

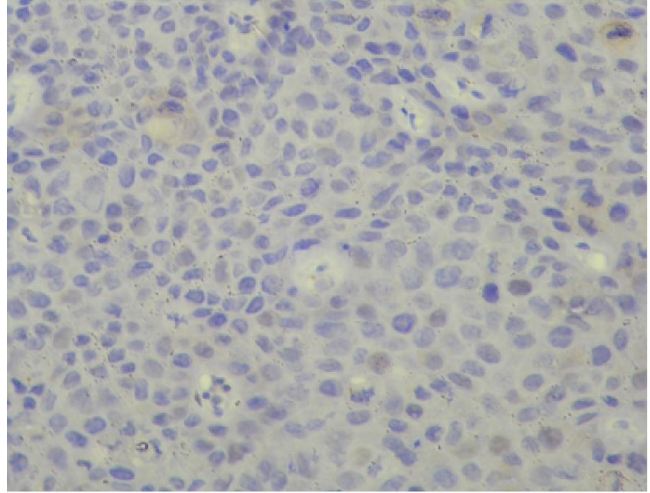


Figure 5. ER expression in SCC

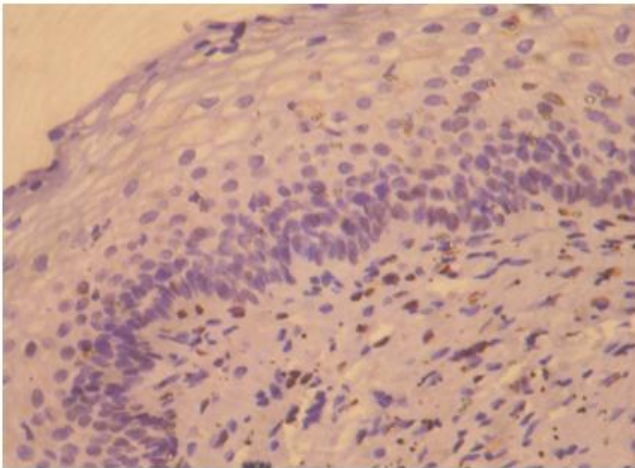


Figure 6. PR expression in normal cervix

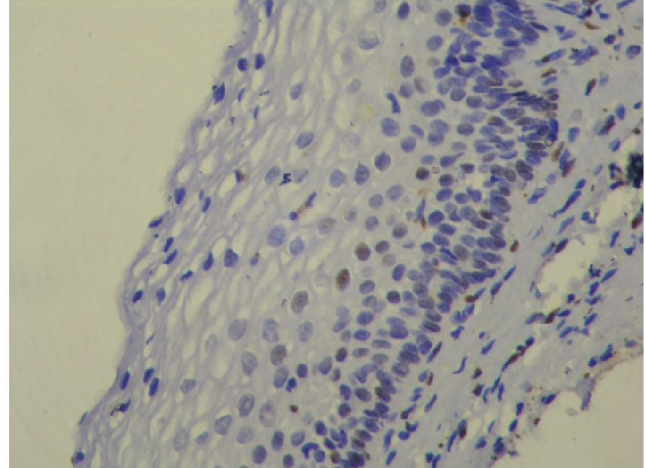


Figure 7. PR expression in LSIL

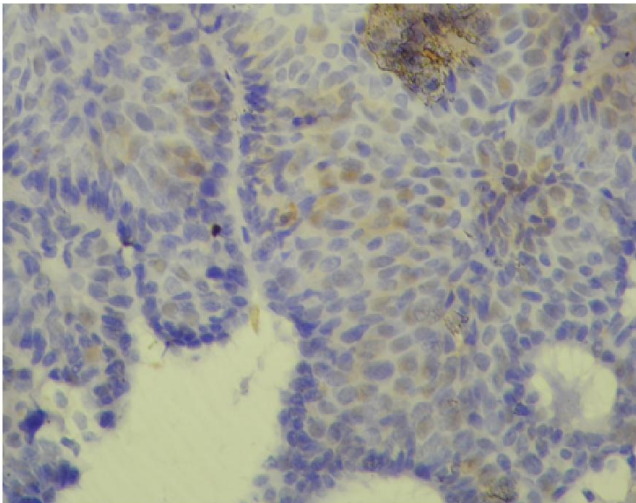


Figure 8. PR expression in HSIL

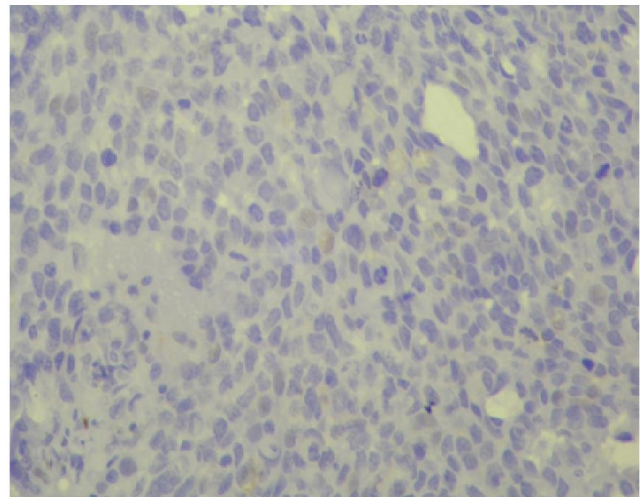


Figure 9. PR expression in SCC

DISCUSSION

Down regulation of ER expression may be the early alteration to take place in normal epithelium during the development of

cervical dysplasia in women infected with HR-HPV (Chung *et al.*, 2010). This indicates a loss of normal growth control by sex steroid hormones (Bekkers *et al.*, 2005). Studies in animal model showed that the ER required for the early stage of

cervical carcinogenesis. It suggests that ER is no longer required once the critical precursor lesion developed (Chung et al., 2008) Transcriptional repression by other transcription factors is likely to contribute to loss of ER during cervical cancer progression (Zhai et al., 2010). PR assembly is an oestrogen-dependent process; the presence of functional ER may be necessary for the formation of PR (Coelho et al., 2004). Our data indicates that there is a loss of ER and PR during the neoplastic transformation. The loss of ER expression was more prominent than that of PR. Loss of ER expression may be associated with malignant transformation in the HR-HPV infected cervical epithelium. Our result can be compared to Kanai et al. (1998) who reported ER and PR expression in normal cervical epithelia and marked reduction of ER and moderate reduction of PR in neoplastic lesions. Our results are contradictory to Kim et al. (1992) who reported increased expression of ER and PR in SCC. In spite of contradictory results in some studies, our study showed a consistent decrease in the expression of ER with increasing degree of dysplasia. This decline in the expression of the receptors may help in early detection and prevention of cervical cancer. Also, the detection of these hormones oestrogen and progesterone in the blood may be a prospective study for early cancer detection. Various other markers like potassium channels, human papilloma virus L1 capsid protein, P-glycoprotein, matrix metalloproteinase 2 and p16(INK4a) are also being investigated as an early marker of cervical cancer (Ortiz et al., 2011, Lee, Sung, 2011, Singh et al., 2012, Lee HJ, Kim JW, 2013, Nankivell et al., 2014). Also traditional Indian herbal molecules like curcumin are studied for their role in suppressing estradiol and reduce the incidence of cervical cancer (Mayank Singh, Neeta Singh, 2011).

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REFERENCES

AngioloGadducci, Cecilia Barsotti, StefaniaCosio, Lavinia Domenici, and Andrea Riccardo Genazzani, 2011. Smoking habit, immune suppression, oral contraceptive use, and hormone replacement therapy use and cervical carcinogenesis: a review of the literature, *Disease Markers.*, 27(8): 597-04.

Bekkers, R. L, Van der Avoort IA, Melchers WJ, Bulten J, de Wilde PC, Massuger LF, Down regulation of oestrogen receptor expression is an early event in human papillomavirus infected cervical dysplasia, 2005. *Eur J GynaecolOncol.*, 26(4): 376-82.

Chung, S. H., MyeongKyun Shin, Kenneth S. Korach, Paul F. Lambert, Requirement for Stromal Oestrogen Receptor Alpha in Cervical Neoplasia, 2013. *Hormones and Cancer*, 4: 50-59.

Chung, S. H., Silvia Franceschi, Paul F. Lambert, Oestrogen and ER α : Culprits in cervical cancer? Review article, 2010. *Trends in Endocrinol. Metab.*, 21(8): 504–11.

Chung SH, Weidmeyer K, Shai A, Korach KS, Lambert PF, Requirement of oestrogen receptor α in a mouse model for human papillomavirus-associated cervical cancer, 2008. *Cancer Res.*, 68(23): 9928-34.

Coelho FR, Prado JC, Pereira Sobrinho JS, Hamada G, Landman G, Pinto CA, Oestrogen and progesterone receptors in human papilloma virus-related cervical neoplasia, 2004. *Braz. J. Med. Biol. Res.*, 37(1): 83-88.

Kanai M, Shiozawa T, Xin L, Nikaido T, Fuji S, Immunohistochemical detection of sex steroid receptors, cyclins, and cyclin-dependent kinases in the normal and neoplastic squamous epithelia of the uterine cervix, 1998. *Cancer*, 82(9): 1709-19.

Kim, J.W., Sung, H.R., Kim, D.K., Song, C.H, Oestrogen and progesterone receptor assay in carcinoma of the cervix with monoclonal antibodies, 1992. *Gynecol. Oncol.*, 47: 306–10.

Lee HJ, Kim JW, Immunohistochemical study on the expression of matrix metalloproteinase 2 and high-risk human papilloma virus in the malignant progression of papillomas, 2013. *J. Korean Association of oral and Maxillofac. Surg.*, 39(5): 224-30.

Lee, Sung, Correlation between immunocytochemistry of human papilloma virus L1 capsid protein and behavior of low-grade cervical cytology in Korean women, 2011. *Journal of Obstetrics & Gynaecology Research*, 37(9): 1222-28.

Mayank Singh, Neeta Singh, Curcumin counteracts the proliferative effect of estradiol and induces apoptosis in cervical cancer cells, 2011. *Molecular and Cellular Biochemistry*, 347: 1-11.

Nankivell P, Williams H, Webster K, Pearson D, High A, MacLennan K, Senguvan B, McConkey C, Rabbitts P, Mehanna H, Investigation of p16(INK4a) as a prognostic biomarker in oral epithelial dysplasia, 2014. *J. Oral Pathol. Med.*, 43(4): 245-49.

Ortiz, Saqui-Salces, Hinojosa, Garcia-Villa, Eagl potassium channels as markers of cervical dysplasia, 2011. *Oncology Reports*, 26(6): 1377-83.

Pearce ST, Jordan VC, The biological role of oestrogen receptors α and β in cancer, 2004. *Crit. Rev. Oncol. Hematol.*, 50(1): 3-22.

Remoue F, Jacobs N, Miot V, Boniver J, Delvenne P, High intraepithelial expression of oestrogen and progesterone receptors in the transformation zone of the uterine cervix, 2003. *Am. J. Obstet. Gynecol.*, 189(6): 1660-65.

Sarpin KM, Graham JD, Mote PA, Clarke CL, Progesterone action in human tissues: regulation by progesterone receptor (PR) isoform expression, nuclear positioning and co-regulator expression, 2009. *Nucl. Recept. Signal*, 7: 1-13.

Singh M, Singh U, Mathur N, Shukla Y, Expression of P-glycoprotein is positively correlated with p53 in human papilloma virus induced squamous intraepithelial lesions of uterine cervix: poor prognosis association, 2012. *Asian. Pac. J. Cancer*, 13(12): 6039-45.

Zhai Y, Bommer GT, Feng Y, Weise AB, Fearon ER, Cho KR, Loss of oestrogen receptor 1 enhances cervical cancer invasion, 2010. *Am. J. Pathol.*, 177(2): 884-95.
