



RESEARCH ARTICLE

EFFECT OF ARSENIC ON CERTAIN BIOCHEMICAL PARAMETERS IN LIVER TISSUE OF *Tilapia mossambica*

Soundararajan, M* and Veeraiyan, G.

Department of Zoology, Annamalai University, Annamalai Nagar-608 002, Tamilnadu, India

ARTICLE INFO

Article History:

Received 9th October, 2010

Received in revised form

13th October, 2010

Accepted 17th November, 2010

Published online 5th December, 2010

Key words:

Biochemical parameters,

Arsenic,

Tilapia mossambica,

Liver.

ABSTRACT

In the present study, the sub-lethal effects of arsenic on various biochemical parameters of *Tilapia mossambica* were studied. The fish was exposed to sub-lethal concentration of arsenic for 15 days for chronic toxicity studies. In the present study, total protein, amino acid and acetylcholinesterase, glycogen and lactic acid were observed. The present study showed the protein content was decreased and amino acid content was increased significantly and also Acetylcholinesterase was increased in the liver tissue of arsenic treated fish, *Tilapia mossambica*. The present study shows the level of glycogen decreased and lactic acid increased in the liver tissue of fish exposed to arsenic. These changes were concentration dependent.

© Copy Right, IJCR, 2010 Academic Journals. All rights reserved.

INTRODUCTION

Arsenic is a toxic element for humans and it is commonly associated with serious health disruptions (Brookes 1998). Total diet as studies carried out in various countries have shown that fish and shell fish are the most significant dietary source of as, accounting for nearly three quarters of total intake (Dokkum *et al.*, 1989, Tao *et al.*, 1999). The concentration of as was found in environmental samples, mainly in waters, where inorganic form is predominant (Smith *et al.*, 2000, Elci *et al.*, 2008). Arsenic exposure has been related to the appearance of some types of cancer (Ranbis *et al.*, 2003). A report on an assessment of the cancer risk associated with consumption of oysters caused a panic among consumers in Taiwan

(Guo, 2002). Some of these human health effects currently observed in population of South and Southeastern Asia, particularly in countries such as Bangladesh and India (AIRamali, 2005). Besides the direct exposure of humans to as through drinking contaminated water, the As might also be biologically available to aquatic organisms, such as fish which are used as human food thereby providing an additional source of as. Arsenic has a considerable tendency to accumulate in bottom sediments (Svobodovo, 2002). For this reason, issues related to As content in aquatic organisms and sea fish in particular, have attracted considerable attentions. The relevance of this as intake will depend on the concentration of As accumulated by the fish (Lai *et al.*, 2001). During recent years, serious concern has been voiced about

*Corresponding author: soundararajan_zoo@yahoo.com

the rapidly deteriorating state of fresh water bodies with respect to toxic metals pollution. Fishes are often at the top of the aquatic food chain and accumulate large amounts of some metals from the water (Tuzen, 2003). Water pollution leads to fish contamination with toxic metals from many sources, e.g., industrial and domestic wastewater, natural runoff and contributory rivers (Rashed, 2001; Tariq *et al.*, 1991). Fishes, living in polluted water may accumulate toxic trace metals via their food chains, they assimilate metals by ingestion of particulate material suspended in water, ion exchange of dissolved metals across lipophilic membranes, e.g., the gills, adsorption on tissue and membranes surfaces (Alam *et al.*, 2002). The bioaccumulation of metals is therefore, an index of the pollution status of the relevant water body (Mastoi *et al.*, 2008). Protein is the most important and abundant biochemical constituent present in the animal body.

Proteins are important in all biological systems. Protein and amino acids are very important nutrients. Protein plays a major role in the synthesis of microsomal detoxifying enzymes and helps to detoxify the toxicants which enter into the animal body (Ramasamy, 1987). Amino acids are the building blocks of protein which are organic compounds, meaning that they contain carbon and hydrogen bonded to each other. In addition to those two elements, they include nitrogen, oxygen, and, in a few cases, sulfur. The basic structure of an amino-acid molecule consists of a carbon atom bonded to an amino group (-NH₂), a carboxyl group (-COOH), a hydrogen atom, and a fourth group that differs from one amino acid to another and often is referred to as the -R group or the side chain.

The -R group, which can vary widely, is responsible for the differences in chemical properties fetus (Sankarsamipillai and Jagadeesan, 2006). Acetylcholinesterase is an enzyme present in various tissues, including muscle and red cells, that breaks down acetylcholine (a chemical released by nerves that activates muscle contractions) and helps to maintain proper transmission of impulses between nerve cells and between nerve cells and muscles; also called true cholinesterase. Measuring acetylcholinesterase in

amniotic fluid may help confirm a suspected neural tube defect in the fetus (Sankarsamipillai and Jagadeesan, 2006). Carbohydrate is an essential energy source for all vital activities of an organism. It is stored in the form of glycogen in animals. Glycogen is broken down into glucose for energy requirements. The stressful condition disturbs the metabolic rate of carbohydrate and thus the level of glycogen, glucose and lactic acid are altered (Srivastava and Singh, 1980; Metevlev *et al.*, 1983). The present study was carried with an aim to investigate the sub-lethal effect of arsenic in biochemical parameters in liver tissue of *Tilapia mossambica*.

MATERIALS AND METHODS

The fresh water fish *Tilapia mossambica* were collected from fish farm at Puthur, Tamil Nadu, India. The collected fish were acclimated to laboratory condition for 15 days. They were checked thoroughly for injury and disease conditions, and only healthy fishes were used for this study. After washing with 0.01% KMnO₄ solution for 15 min, they were placed in nine plastic pools (500 L) containing non-chlorinated water. Prior to the start of the experiment, the fishes were acclimatized to the food and laboratory conditions with 12 h dark and 12 h light cycles, pH range of 6.95 to 7.60 and temperature ranging from 16 to 24 °C for 15 days.

Fishes were divided into three equal groups each comprising of 36 fishes. Each group was kept in separate plastic tanks. The first group was kept as negative control; the fishes were maintained in water containing normal water without any treatment. The fishes of two groups were exposed to a sub-lethal concentration of 1 ppm concentration of Arsenic added in the water for 30 days respectively. Solutions were renewed once daily after exposure period, animals were sacrificed and the liver tissues were removed, homogenized and stored at -80 °C for further biochemical analyses.

Estimation of biochemical study

Protein content in the tissue were estimated by the method of Lowry *et al.* (1951), Total free amino

acids and content of the tissue were estimated by the method of Moore and Stein (1954), The enzyme acetylcholinesterase was assayed by Metcalf (1951), The glycogen content was estimated by Kemp and Kits Van Heijhingen (1954), and Lactic acid was done by the method of Barker and Summerson (1941)

Statistical analysis

The data were subjected to student "t" test to find out the significance of difference between control and treated values.

RESULTS

In the present study, attempts have been made to investigate the effects of sub-lethal concentration of arsenic on various biochemical parameters of *Tilapia mossambica* in acute and toxicity studies. In the liver tissue of control groups, the protein content was 86.92 ± 1.98 mg/g wt. wt. of tissue. After the mercury exposure the level of protein content was significantly decreased in liver tissue of arsenic exposed fish, as compared to respective control levels (Table.1).

Table 1 shows the amino acid content in the brain tissue of fish. The level of protein content was increased in arsenic exposed fish. In the liver tissue of control fish, the acetylcholinesterase activity was 45.72 ± 0.95 μ moles of acetylchoine hydrolysed/mg of protein/hr. During the arsenic exposure the activity of acetylcholinesterase was decreased in the liver tissue of fish (Table.1).

The level of glycogen content in the liver tissue of control fish was 11.99 ± 1.96 mg/g wet wt. of tissue. During the arsenic exposure the level of glycogen decreased in the liver tissue (8.42 ± 0.97 mg/g wet wt. of tissue). In the liver tissue of control groups, The lactic acid content was 2.84 ± 1.08 mg/g wt. wt. of tissue. After the arsenic exposure the level of lactic acid content was significantly decreased in liver tissue of arsenic exposed fish, as compared to respective control levels (Table.1).

DISCUSSION

In the present study, A reduction in the protein content observed in *Tilapia mossambica* exposed with arsenic. These results suggest that the tissue protein undergoes proteolysis results in an increase in the production of free amino acids. These amino acids are utilized for energy production during stressful situation in the intoxicated fishes. Neff (1985) has reported that decline in protein content may also be related to increased energy cost of homeostasis, tissue repair and detoxification during stress. In the present investigation, sublethal concentrations of arsenic exposed fish *Tilapia mossambica* exposed with arsenic show a decrease in protein content and an increase in amino acid content of liver for 15 days exposure of arsenic.

Many investigations have also reported such a change in total protein content of various tissues in different fishes exposed to different heavy metals (Rajamanikam 1992; Pazhanisamy, 2002). Jana and Bandyopathay (1981) have reported such a

Table 1. Level of biochemical parameters in liver tissue of *Tilapia mossambica* treated with arsenic

Parameters	Control	15 days treated
Protein (mg/g wte wt. of tissue)	86.92 ± 1.98	$73.85 \pm 1.68^*$
Amino acid (μ g/g wet wt. of tissue)	2.85 ± 1.54	$3.66 \pm 1.87^*$
Acetylcholinesterase (AchE) (μ moles of acetylchoine hydrolysed / mg of protein/hr)	45.72 ± 0.95	$36.14 \pm 1.92^*$
Glycogen (mg/g wet wt. of tissue)	11.99 ± 1.96	$8.42 \pm 0.97^*$
Lactic acid (mg/g wet wt. of tissue)	2.84 ± 1.08	$4.22 \pm 1.87^*$

Mean \pm S.D of six individual observations
*significance at 5% level

reduction in protein content when the fish *Channa punctatus* has been exposed to heavy metals such as mercury, arsenic and lead. Protein depletion has been reported in the liver of *Anabas testudines* exposed to nickel chloride (Jha and Jha, 1995). Decrease in the liver protein level is reported in the fish *Labeo rohita* exposed to arsenic (Pazhanisamy, 2002), *Channa punctatus* exposed to zinc and phenyl mercuric acetate (Sen *et al.*, 1992; Karuppasamy, 2000) *Channa punctatus* exposed to arsenic (Jatyajit Hota, 1996), *Channa striatus* exposed to mercury cadmium and lead (Palanichamy and Baskaran, 1995) and *Cirrhina mrigala* exposed to lead acetate (Ramalingam *et al.*, 2000). Baskaran *et al.* (1991) have reported the impact of commercial detergent (Nirma) on feeding energetics and protein metabolism in the freshwater teleost fish *Oreochromis mossambicus*.

The decrease in liver and muscle protein has been reported in the sugar mill effluent treated *Channa punctatus* after 96 hr exposure (Avash Maruthi and Ramakrishna Rao, 2000). In the present investigation, the decreased level of protein in brain tissue shows that fish exposed to arsenic are subject to stress. Similar results have also been recorded in the protein content of different tissues when the animals are exposed to various pollutants (Palanichamy *et al.*, 1989; Malla Reddy and Bashamohidran, 1988), Manoharan and Subbiah, 1982).

Meenakshi and Indra (1998) have noticed depletion in the level of total protein in liver and muscle and an increase in the total free amino acids in blood, liver and muscle of distillery effluent treated *Mystus vittatus*. The remarkable increase in the free amino acid may represent proteolysis in the tissues to meet the demands for energy requisites in addition to the carbohydrates and fat. Increase in amino acid content in liver is observed in *Mystus vittatus* exposed to median lethal concentration of mercuric chloride (Jagedeesan, 1994) and in *Mystus vittatus* exposed to sublethal and median lethal concentration of copper (Rajamanickam, 1992). Anuradha and Raju (1996) have observed the increased level of amino acid content in liver, muscle, kidney and gill tissues of *Anabas scandens* exposed to selenium toxicity. The FAA serves as metabolites for a TCA cycle which

have a key role in stepping up the energy requirement. Acetylcholinesterase (AChE) activity measurement in fish has been used for monitoring the neurotoxicity of pesticide (Bretaud *et al.*, 2000). AChE, a serine hydrolase, catalyzes the breakdown of the neurotransmitter acetylcholine into acetate and choline. This process involves the formation of a substrate enzyme complex, followed by acetylation of the hydroxyl group, the amino acid serine, present within the esteratic side and finally deacetylation.

The inhibitory effect on AChE activity indicates that pollutants like insecticide might interfere in the vital processes like energy metabolism of nerve cells (Nath and Kumar, 1999). In the present study, 15 days exposure period of lead has resulted the inhibition of AChE activity level in the brain of *Tilapia mossambica* a decrease in AChE activity level has led to the accumulation of acetylcholine in the brain of fish (Josh *et al.*, 1982). AChE inhibition and an accumulation of ACh in the tissues of sumithion treated fish *Tilapia mossambica* have been observed by Koundinya and Ramamurthi (1978). Bashamohideen and Sailbala (1989) have observed a steep decline in AChE activity with a concomitant elevation in ACh content in different tissues like gill, kidney, brain, liver and different types of muscles in *Cyprinus carpio* following 10 days exposure to malathion. The decrease in brain AChE is found to be inversely proportional to the increase in ACh content in methyl parathion treated tadpoles of frog, *Rana cyanophlicits*. Accumulation of ACh and inhibition of AChE activity levels in liver, muscle, gill and brain have been reported in *Tilapia mossambica* exposed to fenvalerate (Ghosh, 1990). Ravi and Selvarajan (1990) have reported an increase in the levels of amine in the brain region of *Labeo rohita* and *Cyprinus carpio* exposed to phosalone. Sevçiler *et al.* (2004) have reported a significant correlation between increase in lipid peroxidation and inhibition of AChE activity in liver. They have further stated that etoxazole mediated lipid peroxidation may be related to its anticholine esterase action. Increased lipid peroxidation caused by etoxazole indicates that this compound induces the generation of reactive oxygen species, creating oxidative damage in the cell membrane. Yang and Dettab (1996) in their

study with diisopropyl fluorophosphates have suggested that AchE inhibitor induced cholinergic hyperactivity has initiated the accumulation of free radicals leading to lipid peroxidation, which may be the initiator of AchE inhibitor induced cell injury. Nachmanson and Feld (1947) have reported that the animal dies when AchE activity of the brain is inhibited by 95 percent. Coppage *et al.* (1975) have observed 79 percent reduction in AchE activity in the esturine fish *Lagodon rhomboids* exposed to 48 hours median lethal concentrations (92µg/l) of malathion. Quinolphos, another organophosphorus insecticide also produces a highly significant inhibition of AchE activity in the brain and a reduction in RBC of female guinea pigs (Dikshith *et al.*, 1980).

Carbohydrate is an essential energy source for all vital activities of an organism. It is stored in the form of glycogen in animals. Glycogen is broken down into glucose for energy requirements. The stressful condition disturbs the metabolic rate of carbohydrate and thus the level of glycogen, glucose and lactic acid are altered (Srivastava and Singh, 1980; Motelev *et al.*, 1983). The toxic substances are absorbed into the body and transported to various organs through blood. The blood glucose is a sensitive biochemical indicator of stress (Motelev *et al.*, 1983). Exposure of fishes to different types of toxic substance is known to elicit changes in the biochemical constituents and thereby altering the metabolic pathways. In the present study the level of glycogen content and lactic acid was increased in the liver tissue of fish exposed to arsenic. Changes in the glycogen level of liver has been noticed by many investigators. Mcleay and Brown, (1975) have recorded a considerable decrease in glycogen content of bleached kraft pulp mill effluent. Baskaran *et al.*, (1989) have noticed the depletion on the hepatic glycogen content in *Oreochromis mossambicus* when exposed to textile dye effluent. Depletion in the glycogen content of liver and muscle have been observed in *Rasbora daniconirus* exposed to pulp and paper mill effluent (Vijayaran and Vasugi, 1989).

Tilapia mossambica exposed to sublethal concentration of arsenic shows an overall increase in the blood glucose at all periods of exposure,

thereby indicating that the glycogenolysis takes place in the liver, where by the reserved glycogen is being slowly converted into glucose. The hyperglycemic condition in the present study correlated with the observations of some researcher's *viz.*, the juvenile Cohosalmon on *Corhynchus kisutch* treated with sub-lethal concentration of neutralized unbleached kraft mill effluent (Mcleay, 1973). Similar results were made by Vijayram, and Vasugi, (1989) in paper and pulp mill effluents. Similar elevated blood glucose levels have been noticed in *Meteropheustes fossilis* exposed to textile mill effluent (Nisha and Shukla, 1986).

Lactic acid is formed through glycolysis under anaerobic condition of glucose catabolism. In the present study *Cyprinus carpio* showed an increase in the lactic acid content of liver and blood at all the hours of effluent treatment. Accumulation of lactic acid is more in liver and blood of fishes exposed to raw effluent. It is likely that the lactic acid formed in the muscle and other tissue during glycolysis, might have been transported to liver *via* blood accounting for the hyper lactamia in blood and liver. Because of the absence of the enzyme glucose - 6 phosphatase in the muscle, which is necessary for the conversion of lactic acid into glucose, the lactic acid produced in the tissue is transported to the liver through blood (Ambika shanmugam, 1980). Since liver is the metabolic site, the lactic acid transported from the tissue to liver is utilised for the resynthesis the of glucose and glycogen through cori cycle (Mayer Bodensky, 1947) contributing to the increase in the level of lactic acid in liver and blood at all periods of study. Burton *et al.*, (1972) have observed the heavy accumulation of lactic acid in liver of rain bow trout *Salmo gairdneri* exposed to zinc.

Acknowledgement

The authors are grateful to Professor and Head, Department of Zoology, Annamalai University to complete the work successfully.

REFERENCES

- Alam, M.G.M., Tanaka, A., Allinson, G., Laurenson, L.J.B., Stagnitti, F. and Snow, E.T.A. 2002. Comparison of trace element

- concentrations in cultured and wild carp (*Cyprinus carpio*) of lake Kasumigaura. *Jpn. Ecotoxicol. Environ.*, 53: 348–354.
- AIRmali, S.W., Haris, P.I., Harrington, C.F., and Ayub, M.A. 2005. Survey of arsenic in food stuffs sale in the United Kingdom and imported from Bangladesh. *Sci. Total. Environ.*, 337: 23–30.
- Ambika shanmugam, 1980. Fundamentals of Biochemistry for medical students. Novabharat offset Work pp. 350-411.
- Anuradha, C.H., and Raju, T.N. 1996. Effect of selenium toxicity on nuclei acid, protein and free amino acid contents of the fish *Anabas scandens*. *J. Ecotoxicol. Anabas scandens J. Ecotoxicol. Environ. Monit.*, 6(3):163-166.
- Avasan Maruthi, Y., and Ramakrishna Rao, S. 2000. Effect of sugar mill effluent on organic reserves of fish. *Poll. Res.*, 19(3): 391-393.
- Barker S.B and Summerson, W.H. 1941. The colorimetric determination of lactic acid in biological material. *J. Biol. Chem.*, 138 : 535 - 554.
- Bashamohideen, M. and Saibala, T. 1989. Acetylcholinesterase activity in the tissues of the common carp *Cyprinus carpio* (Linnaeus) subjected to the sub-lethal exposure of malathion. *J. of Environ. Biol.*, 10(1): 51-57.
- Bashamohideen, M. and Saibala, T. 1989. Acetylcholinesterase activity in the tissues of the common carp *Cyprinus carpio* (Linnaeus) subjected to the sub-lethal exposure of malathion. *J. of Environ. Biol.*, 10(1): 51-57.
- Baskaran, P., Palanichamy, S. and Arunachalam, S. 1989. Effects of textile dye effluent on feeding energetic, body composition and oxygen uptake of the freshwater fish *Oreochromis mossambicus*. *J. Ecobiol.*, 1: 203-214.
- Bretau, S. J., Toutant, P. and Saglio, P. 2000. Effects of carboduran diuron and nicosulfuron on acetylcholinesterase activity in goldfish (*Carassius auratus*), *ecotoxicol. Environ. Saf.*, 47. 117-124.
- Brookes, R.R. 1998. Plants that hyper accumulate heavy metals, in: Their Role in Phyto remediation, microbiology, Archaeology, Mineral Exploration and Phytomining, CAB International, Wallingford, UK.
- Burton, T.D., Jones, A.H. and Cairns, J.J. 1972. Acute zinc toxicity to rainbow trout (*Salmo gairden*) confirmation of the hypothesis that death is related to tissue hypoxia. *Fish. Res. Bd. Can.*, 29: 1463-1466.
- Coppage, D.L., 1972. Organophosphate pesticides : Specific level of brain AchE inhibition related to death in sheep head minnow. *Trans. Ameri. Fish. Soc.*, 101, 534.
- Dikshith, T.S.S., Raizda, R.B. and Datta, K.K. 1980. Response of female guinea pigs to repeated oral administration of quinolphos. *Bull. Enviorn. Contam. Toxicol.*, 24: 739-745.
- Dokkun, W.V., Devos, R.H., Muys, T.H. and Wesstra, J.A. 1989. Minerals and trace elements n total diets in the Netherlands. *Br. J. Nutr.*, 61: 7–15.
- Elci, L.U. Divrikli, M. and Soylak, 2008. Inorganic arsenic speciation in various water samples ith GF-AAS using coprecipitation. *Int. J. Environ. Anal. Chem.*, 88.711–723.
- Ghosh, T.K., 1990. Synthetic pyrethroid intoxication on tissue acetylcholine and acetylcholinesterase in the fish, *Tilapia mossambica*. *Environ. Ecol.*, 8(3): 950-954.
- Guo, H.R. 2002. Cancer risk assessment for arsenic exposure through oyster consumption. *Environ. Health Perspect.*, 110:123–124.
- Jana, S. and Bondyopadhyay, N.1981. Effect of heavy metals on some biochemical parameters in the freshwater fish, *Channa punctatus*. *Environ. Ecol.*, 5(3): 488-493.
- Jatyajit Hota, 1996. Arsenic toxicity to the brain, liver and intestine on a freshwater fish, *Chnna punctatus* (Bloch). *Geobios*, 23 :154-156.
- Jha, B.S and Jha, M.M. 1995. Biochemical effects of nickel chloride on the liver and gonads of the freshwater climbing perch, *Anabas testudineus* (boch). *Proc. Nat. Acad. Sci. India*, 65(B): 39-46.
- Karupphasamy, R. 2000. Short and long term effects phenyl mercuric acetate on protein metabolism in *Channa punctatus* (Bloch). *J. Natcon.*, 12(1): 83-93.
- Kaundinya, R.P. and Ramamurthy, R. 1978. Effect of sumithion (Fentothion) on some selected enzyme system in the fish, *Tilapia mossambica* (peters). *Indian J. Exp. Biol.*, 16: 809-811.
- Kemp, A and Kitsven Hejhingeen, J.M. 1954. A colorimetric micro method for the

- determination of glycogen in tissues. *Biochem. J.*, 56: 640-648.
- Lai, W.M.W. Cullen, S. Ray, 2001. Arsenic speciation in sea scallop gonads, Appl. Lake in Sindh (Pakistan), *Environ. Monit. Assess.* 141 (2008) 287-296.
- Lowry, O.H., Rosenbrough, N.J., Farr, A. and Randall, R.J. 1951. Protein measurement with folinphenol reagent. *J. Biol. Chem.*, 193: 265-273.
- Jakubis, M. and Nieuwenhuijsen, M.J. 2003. EXPASCAN Study Group, Association between arsenic exposure from a coal-burning power plant and urinary arsenic concentrations in Prievidza District, Slovakia. *Environ. Health Perspect.*, 111: 89-894.
- Malla Reddy, P. and Basha Mohideen M.D. 1988. Toxic impact of fenvalerate on the protein metabolism in the branchial tissue of a fish *Cyprinus carpio*. *Curr. Sci.*, 57: 211-212.
- Manoharan, T. and Subbiah, G.N. 1982. Toxic sublethal effect of endosulfan on *Barbus stigma*. *Proc. Indi. Acad. Sci. Anim. Sci.*, 91(6): 523-532.
- Mayer Bodensky. 1947. Introduction to physical chemistry. John Willey and Sons, New York, 314-462.
- McLeay, D.J. 1973. Effects of a 12 hour and 25-days exposure to kraft pulpmill effluent on the blood and tissues of juvenile coho salmon (*Oncorhynchus kisutch*). *J.Fish. Res. Bd. Can.*, 30: 395-400.
- McLeay, D.J. and Brown, D.A. 1974. Growth and stimulation and biochemical changes in juvenile *Coho salmon (Oncorhynchus kisutch)* exposed to bleached kraft pulpmill effluent for 200 days. *J.Fish. Res. Bd. Canada.*, 31: 1043-1049.
- Meenakshi, V. and Indra, N. 1998. Sublethal toxicity of distillery effluent on the protein and free aminoacids of the freshwater fish, *Mystus vittatus* (Bloch). *J. Natcon.*, 10(1): 87-91.
- Metcalf, R.L. 1951. Methods in biochemical analysis. (Glick, D. Eds.) Vol.V. Interscience publication. New York.44.
- Meteliev, V.V., Kanaev, A.I. and Dzasokhova, N.G. 1983. Water Toxicology. Amerind Publishing Co. Pvt. Ltd., New Delhi. pp. 56-60.
- Nachmanson, D. and Feld, F.A 1974. In : Insecticides action and metabolism (Ed., O' Brien, R.D). Academic Press, Inc., New York.
- Nath, B.S. and Kumar, R.P.S. 1999. Toxic impact of organophosphorous insecticides on acetylcholinesterase activity in the silkworm, *bombyxmori L. Ecotoxicol, Environ. Saf.*, 42 157-162.
- Neff, J.M. 1985. Use of biochemical measurement to defect pollutant mediated damage to fish. *ASTM. Spec. Tech. Publ.*, 854: 155-183.
- Nisha and Shukla, N.P.1986. Effect of textile liquid effluent on the freshwater fish. *Indian J. Environ. Prot.*, 69(3): 189-192.
- Palanichamy, S., Arunachalam, S. and Baskaran, P. 1989. Impact of pesticides on protein metabolism in the freshwater catfish, *Mystus vittatus*. *J. Ecobiol.*, 1: 90-97.
- Palanisamy, S. and Baskaran, P. 1995. Selected biochemical and physiological response of the fish *Channa striatus* as biomonitor to assess heavy metal pollution in freshwater environment. *J. Ecotoxicol. Environ. Monit.*, 5(2): 131-138.
- Rajamanickam, C. 1992. Effects of heavy metal copper on the biochemical contents, bioaccumulation and histology of the selected organs in the freshwater fish, *Mystus vittatus* (Bloch) Ph.D Thesis, Annamalai University.
- Ramalingam V., Vimaladevi, V., Narmadaraj, R. and Prabakaran, P. 2000. Effect of lead on hematological and biochemical changes in freshwater fish, *Cirrhina mrigala pull. Rs.*, 19(1): 81-84.
- Ramasamy, M. 1987. Effect of sevin on blood free amino acids level of the fish *Sarotherodon mossambicus*. *Environ. Ecol.*, 5: 633-637.
- Rashed, M.N. 2001. onitoring of environmental heavy metals in fish from Nasser Lake. *Environ. Int.*, 27. 27-33.
- Sankar Sampillai, S. and Jagadeesan, G. 2006a. Protective role of taurine on mercuric chloride induced neurotoxicity in rats. *Poll. Res.*, 25: 39-43.
- Sen, G., Bhera, M.K. and Patel, P. 1992. Effect of zinc on haematobiochemical parameters of *Channa punctatus*. *J. Ecotoxicol. Environ. Monit.*, 2(2): 89-92.
- Sevgiler, Y., Elif OzCan Oruc, and Nevin Uner (2004). Evaluation of etoxazole toxicity in the

- liver of *Oreochromis niloticus* pesticide biochemistry and physiology 78 : 1-8.
- Smith, W.O., Marra, J., Hiscock, M.R. and Barber, R.T. 2000. The seasonal cycle of phytoplankton biomass and primary productivity in the Ross Sea, Antarctica, Deep-Sea Res. PII 47: 3119–3140.
- Srivastava, A.K. and Singh, N.N. 1980 Observation of hyperglycemia in the murrel *Channa punctatus* after acute exposure to methyl parathion. *Comp. Physiol. Ecol.*, 5: 100.
- Svobodovo, Z.O. Celechovska, T. Randak, J. and Machova, 2002. Content of arsenic in arket-ready rainbow trout (*Oncorhynchus mykiss*). *Acta. Vet. Brno.*, 71.361–367.
- Tao, S.S., Bolger, P.M. and Gosh, G.G.D. 1999. Dietary arsenic intakes in the United States. FDA Total Diet Study, September 1991 to December 1996, *Food. Addit. Contam.* 16 465–472.
- Tariq, J.M. Jaffar, M. and Moazzam. 1991. Concentration correlations between major cations and heavy metals in fish from the Arabian Sea, *Mar. Pollut. Bull.*, 22:562–565.
- Tuzen, M. 2003. Determination of heavy metals in fish samples of the middle Black Sea (Turkey) by graphite furnace atomic absorption spectrometry. *Food Chem.*, 80:119–123.
- Vijayram, K. and Vasugi, S.R. 1989. Sublethal effects of pulp and paper mill effluents on the biochemistry of a freshwater fish *Rasbora daniconius*. *Indian J. Environ. Hlth.*, 31(1) : 36-42.
- Yang, W.D. Dettbarn, 1996. Diisopropylphosphorofluoridate-induced cholinergic hyperactivity and lipid peroxidation. *Toxicol. Appl. Pharmacol.*, 138 :48-53.
